Pathology Section

Histomorphological Spectrum of Duodenal Pathology in Functional Dyspepsia Patients

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ABSTRACT

Introduction: Functional Dyspepsia (FD) is one of the most common causes of gastrointestinal symptoms aetiology of which is poorly understood.

Aim: To study duodenal histomorphological features and their relationship with *Helicobacter* pylori (*H Pylori*) infection in patients of FD.

Materials and Methods: This case control study included 50 cases of FD patients selected according to Rome III criteria and 30 age and sex matched controls. These were subjected to oesophago-gastro-duodenoscopy, rapid urease test for detection of *H. pylori* on gastric antral biopsy and duodenal biopsy from second part of duodenum for histopathological evaluation by light microscopy. Ten antral urease positive cases of FD with highest Intraepithelial Lymphocyte Count (IEL) were subjected to Immunohistochemistry (IHC).

Results: Duodenal inflammation was an invariable feature noted in FD. Morphological spectrum consisted of increased IEL in 72%, increased duodenal eosinophils in 92%, presence of focal villous atrophy in 16%, lymphoid aggregates, colonic metaplasia, and duodenal *H. pylori* infection in 4% each. Gastric *H. pylori* positivity was noted in 48% cases of FD. Increased duodenal IEL count and duodenal eosinophilia was noted in 75%, 87.5% such cases. Same was noted respectively, with 61.5% and 95.15% cases with gastric *H. pylori* negativity. In cases of FD, duodenal IEL and eosinophil count in lamina propria showed statistically significant rise when compared with control and had positive correlation with gastric *H pylori* infection. On IHC, increased expression of CD 8 was noted in duodenal IEL and lymphocytes in lamina propria as compared to CD4.

Conclusion: Our study provided some insight in pathogenesis of FD and role of *H. pylori* in its aetiology.

Keywords: Duodenal eosinophils, Inflammation, Intraepithelial lymphocytes

INTRODUCTION

FD is one of the most common causes of gastrointestinal discomfort. It is defined by Rome III criteria as syndrome with one or more of following symptoms which include bothersome postprandial fullness, early satiation, epigastric pain and burning with no evidence of structural disease as seen in upper endoscopy that is likely to explain the symptoms. These criteria should be fulfilled for at least three months with symptom onset, at least six months previously [1].

Aetiology of FD is poorly understood. Local factors like delayed gastric emptying, impaired proximal gastric accommodation to food, gastric hypersensitivity with decreased pain threshold during stomach distension are blamed for the same [2,3]. Activation of immune system with release of mediators like cytokines, nitric oxide, histamine and protease which interfere with function of enteric nerves is the another proposal [4]. According to some studies *H Pylori* infection can be responsible for FD not only by virtue of bringing out gastric but duodenal inflammation as well [5]. Much is being said about duodenal epithelial function, mucosal defense barrier and appropriate signaling systems in patients with FD [6]. Potential role of duodenal IEL and eosinophils in lamina propria has also been described in relation to FD [7,8].

In view of this, we thought that study of histomorphological spectrum of duodenal mucosal pathology with special reference to IEL and eosinophils in lamina propria will be worthwhile in evaluating cases of FD.

MATERIALS AND METHODS

Study design: This prospective study was performed at a Tertiary Care Hospital (BJGMC and SGH Pune, Maharashtra, India) for a period of two years from January 2011 to December 2012. This case control study comprised of 50 patients of FD defined by Rome

III criteria and 30 age and sex matched controls. The controls comprised of patients in whom oesophago-gastro-duodenoscopy (OGDscopy) was done for indications other than FD. After seeking Ethics Committee approval and informed consent, both patients and controls were subjected to OGDscopy. Patients with history of peptic ulcer disease, gastrointestinal malignancy, previous gastric surgery, drug intake and upper gastrointestinal bleeding were excluded from the study.

The study group was evaluated for socio demographic variables (e.g., age, sex), dietary habits, drug history (e.g., NSAIDs, antacids) and gastrointestinal symptomatology. OGDscopy was done and findings were noted. Two biopsies were taken, one each from gastric antrum and second part of duodenum. Rapid urease test was performed for *H. pylori* status on gastric biopsies in gastroscopy room using CLO test kit. A positive result was indicated by colour change from yellow to pink. Duodenal biopsies were fixed in 10% formalin and processed routinely. The sections were cut at 5 micron using rotary microtome and stained with H & E stain, Geimsa and Methylene blue stain.

Detailed histopathological examination of duodenal biopsies was done for grading of inflammation and quantification of duodenal IEL and eosinophils in lamina propria as described below.

Grade of inflammation: Duodenal inflammation was graded by two methods. The subjective grading (as mild, moderate and severe inflammation) was based on overall impression of density of infiltrate in lamina propria and villous morphology. Second is objective grading.

While grading inflammation objectively we used following criteria: density of infiltrate in lamina propria when compared with that in control, counts of IEL and eosinophils in lamina propria and villous architecture [Table/Fig-1]. The findings of metaplasia and presence

of *H. pylori* in duodenum were noted separately. Scoring system for objective grading of duodenal inflammation has been given in [Table/Fig-1].

On summation of individual scores, score of 1 to 4 was graded as mild inflammation, 5 to 8 as moderate inflammation and score more than (>) 8 as severe inflammation.

Infiltrate in lamina propria	IEL count / 100 Entero- cytes	Eosinophil count/ 5 HPF	Villous archi- tecture	Score for each pa- rameter
Within normal limits	Up to 18	Up to15	Normal	0
Overall increased density of cells	19-30	16-30	Focal blunting	1
Overall increased density of cells + presence of polymorphs	31- 40	31-40	Diffuse blunting	2
Overall increased density of cells + Lymphoid follicle or dense focal collection	>41	41-50	Focal/Diffuse blunting	3
Overall increased density of cells + presence of polymorphs +Lymphoid follicle or dense focal collection of lymphocyte	>50	>50	Focal/Diffuse blunting	4 each

[Table/Fig-1]: Scoring system for objective grading of duodenal inflammation.

IEL were counted per 100 enterocytes lining the villous or by villous tip method. Eosinophils were counted in five consecutive non overlapping High Power Fields (HPF) in lamina propria.

Ten cases with IEL count higher than 27/100 enterocytes were further evaluated with IHC for CD4 and CD8 typing of lymphocytes.

STATISTICAL ANALYSIS

The data collected was analysed using SPSS-17.0 software. Chi-Square test was applied to find out significance of difference between means. Correlation t-test and logistic regression analysis was used to find out the correlation between different parameters. The p-value of < 0.05 was considered to be significant.

RESULTS

Age and Sex distribution: The mean age for control was 36.3 years with age range of 20-51. The mean age for FD cases was 34.2 years with age range of 20-63 years. There was slight male preponderance in group of FD with 32 male patients and 18 female patients.

Gastric *H. pylori* **status based on rapid urease test:** Rapid urease test was positive in 24 (48%) cases of FD. Staining of H *pylori* was better with methylene blue stain than with Giemsa stain.

Duodenal Inflammation: When biopsies were examined subjectively in patients of FD, it was possible to rate them as mild inflammation in 23 cases (46%), moderate inflammation in 24 cases (48%) and severe in three cases (6%). Grading of inflammation changed as follows when we resorted to objective grading system. It was mild grade for 31 cases (62%) and moderate grade for 19 (38%) cases in FD.

Villous atrophy: Variable villous abnormality, the form of focal changes of mild flattening of villi was noted in eight cases. Of these, seven were associated with gastric *H. pylori* infection. The range of IEL was from 12 to 37/100 enterocytes with mean of 25.62. The same for eosinophil count/5 high power field was 12 to 39 with mean of 24.5. One case each with gastric *H. pylori* positivity and negativity had normal values for IEL. One gastric *H. pylori* positive case showed normal eosinophilic count in duodenum. Two cases

with gastric *H. pylori* infection were associated with moderate degree inflammation. One of these had goblet cell metaplasia and presence of lymphoid follicles in mucosa in addition and highest values of eosinophils in the group. The other case had highest value of IEL in the group. Closer look at the observation of cases of villous atrophy suggest that, two cases of variable villous abnormality were seen with food allergy with or without gastric *H. pylori* infection while rest six cases were associated with *H. pylori* infection.

Lymphoid aggregates: It was noted in two cases and both these cases had variable villous abnormality. The lymphoid follicles were seen in areas away from variable villous abnormality (focal villous atrophy). Both these cases were positive for antral *H. Pylori*. One case had mild inflammation while the other had moderate inflammation.

Metaplasia: Colonic type of goblet cell metaplasia was noted in two cases. Both showed antral *H. pylori* positivity and moderate degree inflammation. In one case it was associated with variable villous abnormality and lymphoid follicles. The counts for IEL in these cases were 30 each and that for eosinophils 39 and 88 each.

H. Pylori in duodenum: Presence of few *H. pylori* organisms was noted in two cases.

Intraepithelial Lymphocytes: The mean IEL counts for control was 9.3 ± 2.5 with range of 5-18/100 enterocytes, while that of FD cases was 26.18 ± 5.9 with range of 11-37/100 enterocytes. About 70% cases (n=35) showed IEL count to be in the range of 21-30 lymphocytes / 100 enterocytes [Table/Fig-2].

IEL count was increased in 36 (72%) cases of FD when compared to higher limit of our own controls. Of these 20 cases (55.55%) were rapid urease positive. The difference between mean IEL count of H. pylori positive cases and controls was statistically significant with p-value <0.001, while that between H. pylori negative cases and controls was not statistically significant.

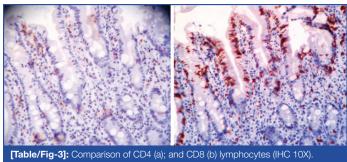
On immunohistochemical evaluation of ten antral rapid urease positive cases with increased IEL counts (> 27/100 enterocytes), all showed increased CD8+lymphocytes compared to CD4+lymphocytes both in intraepithelial location and in lamina propria [Table/Fig-3].

Eosinophil count: Mean eosinophil count was 9.5 ± 2.04 for controls with range of 6-15/ HPF and that of patients with FD was 40.7 ± 26.9 with range of 6 to 134/ 5 HPF. In FD, maximum observations (n=19) were in the range of 21 to 30 eosinophils/5 HPF [Table/Fig-4].

When compared with highest values of our own controls, 92% (n=46) patients with FD showed duodenal eosinophilia. Eosinophilia was seen in 21 gastric rapid urease positive cases and 25 out of 26 rapid urease test negative cases. The difference between mean eosinophilic count of *H. pylori* positive cases and controls was statistically significant with p-value 0.012.

Gastric H. Pylori status	Status of IEL and Ec	Inference	
Rapid Urease test positive n= 24	Raised IEL and Eosinophil Count Raised IEL and normal Eosinophil count	36% (n=18) 4% (n= 02)	H. pylori infection
	Normal IEL and raised Eosinophil Count Normal IEL and Eosinophil count	6% (n= 03) 2% (n= 01)	H. pylori infection /???Food Allergy ?Motor dysfunction
Rapid Urease test Negative n = 26	Raised IEL and Eosinophil Count	32% (n=16)	Other infections / Food Allergy
	Normal IEL and raised Eosinophil Count	18% (n= 09)	Food Allergy
	Normal IEL and Eosinophil Count	2% (n= 01)	?Motor dysfunction

[Table/Fig-2]: Overall status of H. Pylori, IEL and eosinophils.



[Table/Fig-4]: Intraepithelial lymphocytes (H & E, 40X). [Table/Fig-5]: Eosinophils in lamina propria (H & E, 40X). (Images left to right)

Correlation of variables of duodenal inflammation:

IEL correlated positively with presence of gastric *H. pylori* with Chisquare value of 11.45 and p-value of <0.001 with Cl range of 2.2 to 28.7. This wide range of Cl can be attributed to moderate sample size

Duodenal eosinophil count showed positive correlation with positive gastric *H. pylori* status with Chi-square value of 6.2 and p-value of 0.012 with Cl range of 1.59 to 3.04.

On attempting linear regression analysis of different morphological variables of duodenal biopsy with enter method, only IEL showed positive correlation with variables like grade of inflammation, eosinophil count in lamina propria, lymphoid aggregates, metaplasia and villous atrophy. The range of CI however was wide with values of 2.15 to 41.75 perhaps due to moderate sample size.

DISCUSSION

FD is a heterogeneous disorder frequently seen in general population. It's prevalence in India is 13.4% [9]. Present study was conducted to find out possible immune mechanism underlining FD. This was done by quantifying and characterizing duodenal intraepithelial lymphocytes both in *H. pylori* negative and positive patients. In our study, we also tried to find out role of eosinophils in FD.

Mild to moderate degree duodenal inflammation was an invariable feature noted in our patients of FD suggesting its role in symptomatology. Pathogenesis and significance of duodenitis in FD is being elucidated recently. It is suggested that gut-brain-microbial axis plays an important role in functional gut disorders [10]. Recent studies also show persistence of duodenal inflammation in post infection FD [11]. Traditionally stomach pathology is believed to be responsible for dyspeptic symptoms. Recent studies however suggest that the site of pathology may instead be duodenum. Abnormally raised intra epithelial lymphocytes and eosinophils are frequently noted in duodenum in *H. pylori* dyspepsia [12]. Suzuki H et al., have stated that *H. pylori* infection evokes inflammation not only in gastric mucosa and muscularis but also in duodenal mucosa [13].

According to some studies, the persistence of inflammation results from impaired ability of immune system to terminate the inflammation after acute infection by *H. pylori*, *Salmonella* or *Giardia* organisms. IEL by virtue of their strategic location play a great role in preservation of mucosal integrity and maintenance of tolerance to oral particulate or soluble antigens [14].

Hayat M et al., have noted raised IEL in second part of duodenum in 30% cases of *H. pylori* gastritis which persisted even after *H. pylori* were eradicated from gastric location [15].

Raised duodenal IEL [Table/Fig-2] was a feature noted in 72% of our cases of FD. In gastric *H. pylori* positive group, it was seen in 83.33% cases. Experience is shared by others. Lorenzo M et al., in addition demonstrated that majority of duodenal IEL to be CD8+, TIA-1+ and Granzyme B+ which is considered to be activated cytotoxic phenotype [7,16]. Incidentally the duodenal IEL in present study showed markedly raised expression of CD8+ when compared with CD4. This suggests activation of CD8+ lymphocytes without priming with CD4+ cells.

Triad of villous abnormality, raised IEL and chronic duodenal inflammation has been referred as "celiac" lesion in literature. Numerous aetiological agents can elicit such tissue response which mimics celiac disease [17-19]. Such lesions were seen in six of our *H. pylori* positive cases of FD. On clinical grounds we have ruled out possibility of latent celiac disease. These cases can be attributed to *H. pylori* infection.

Closer look at the observation of cases of villous atrophy suggest that, two cases of variable villous abnormality were seen with food allergy one of which had gastric *H. pylori* infection.

More than 70% of Indian population is chronically infected by *H. pylori* [20]. Prevalence of *H. pylori* in present series of FD was 48%. Same in a study from Kashmir has been shown to be 58% [21]. We found two cases of goblet cell metaplasia in duodenum in our gastric *H. pylori* positive patients of FD. Goblet cells can express both antigens of gastric and intestinal mucosa and are considered to represent local precursors of gastric metaplasia [20].

Normal value of IEL in duodenum is subject of individual variability. In present study, highest values of control were 18/100 epithelial cells. Hayat M et al., found it be less than 20/100 epithelial cells [15]. The count of 25 is considered as pathological, between 25 to 30 as border line and that of 30 and more as overtly pathological. Findings in present study are in accordance with this view. The IEL counts of 30 and above were seen in seven cases, between 25 to 30 in 17 cases, between 20 to 24 in 11 cases and a count of 19 in one case of FD.

Very interestingly a spectacular finding of almost invariable duodenal eosinophilia [Table/Fig-4] was noted in our patients of FD (92%). Eosinophils which have different effect or functions are very aptly considered biomarker of FD. They play a great role in allergic inflammation, in host defence mechanism against helminths and in dealing with bacterial and viral infections. H. pylori infection influences duodenal eosinophils. Raised duodenal eosinophils are noted in patients of FD and are suggested to be related to early satiety. It is now almost well established that eosinophils and mast cells are key components of gut hypersensitivity disorder or functional gastrointestinal disorders [22, 23]. FD and Irritable Bowel Syndrome (IBS) are functional disorders and have overlapping symptoms. IEL are of no help in discriminating them but duodenal eosinophils can do this job. While duodenal eosinophilia is linked with FD, mast cell hyperplasia is a feature of IBS [24]. Incidentally both these counts are increased in children with FD triggered by food allergen [25].

Walker MM et al., have suggested that eosinophil-mast cell-neural pathway is involved in functional gastrointestinal disorders [24]. It is a hypersensitivity reaction triggered by pathogen, food or other allergen in which eosinophils or mast cells can up regulate serotonin release which modulates enteric and central nervous system and release lipid mediators and leucotriens. These are very potent stimulators for smooth muscle contraction and thereby responsible for dyspeptic symptoms like abdominal pain and bloating.

Some authors have found a significant correlation between the appearances of symptoms of food hypersensitivity and the presence of IgE-bearing cells and activated eosinophils in gastrointestinal tract. These patients exhibited negative skin prick test and absence of IgE antibodies to the offending food. The gastrointestinal symptoms seen in these patients can be attributed to a localized IgE-mediated response [26].

Considering all these experiences, however simplistic it may feel we have attributed finding of raised duodenal IEL and or eosinophilia in gastric *H. pylori* positive patients to *H. pylori* infection, that with raised IEL in gastric *H. pylori* negative group to some other infection, that of normal IEL and raised eosinophil in gastric *H. pylori* negative patients to food allergy and that with normal IEL and eosinophil to pure motor dysfunction as shown in [Table/Fig-2]. Decreased clearance and increased exposure to duodenal lipids or acid due to reduced duodenal motor response and altered duodeno- jejunal motility are some of the motor dysfunction factors explaining FD symptoms [27,28]. We had two such cases which had normal IEL and eosinophil count. One of such cases was *H. pylori* positive and the other was *H.pylori* negative. The findings are summarized in [Table/Fig-5].

It will not be too pragmatic to state that screening and eradication of *H. pylori* along with anti allergic treatment should be a primary concern in treating patients with FD. A simple measure of small and frequent meals will also go long way in treating patients of FD.

LIMITATION

Moderate sample size was the main limitation of the study along with lack of definitive evidence to support the role of food allergy and motor dysfunction in aetiology of FD.

CONCLUSION

This comprehensive study of duodenal morphology provided some insight in pathogenesis of FD and role of *H. pylori* in it. It definitely makes sense in screening and eradicating *H. pylori* in patients with FD.

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REFERENCES

- Tack J, Talley NJ, Camilleri M, Holtmann G, Hu P, Malagelada JR. Functional gastroduodenal disorders. Gastroenterology. 2006;130:1466-79.
- [2] Pallotta N, Pezzotti P, Corazziari E. Relationship between antral distension and postprandial symptoms in functional dyspepsia. World J Gastroenterol. 2006;12:6982-91.
- [3] Choung RS, Talley NJ. Novel mechanisms in functional dyspepsia. World J Gastroenterol. 2006;12:673-77.
- [4] Delvaux M. Alterations of sensory-motor functions of the digestive tract in the pathophysiology of irritable Bowel syndrome. Best Pract Res Clin Gastroenterol. 2004;18:747-71.
- [5] Fan X, Crowe SE, Behar S, Gunasena H, Ye G, Haeberle H, et al. The effect of class II MHC expression on adherence of *Helicobacter pylori* and induction of apoptosis in gastric epithelial cells: A mechanism for Th1 cell-mediated damage. J Exp Med. 1998;187(10):1659-69.

- [6] Witte AB, D'Amato M, Poulsen SS, Laurent A, Knuhtsen S, Bindslev N, et al. Duodenal epithelial transport in functional dyspepsia: Role of serotonin. World J Gastrointest pathophysiol. 2013;4(2):28-36.
- [7] Gargala G, Lecleire S, François A. Duodenal intraepithelial T lymphocytes in patients with functional dyspepsia. World J Gastroenterol. 2007;13:2333–38.
- [8] Walker MM, Talley NJ. Functional gastrointestinal disorders and the potential role of eosinophils. Gastroenterol Clin North Am. 2008;37:383–95.
- [9] Ghoshal UC, Singh R. Functional dyspepsia: The Indian scenario. JAPI. 2012;60:6-7.
- [10] Lee YY, Chua AS. Influence of gut microbes on the brain gut axis. (Gut. 2011;60:307-317). J Neurogastroenterol Motil. 2011;17:427-29.
- [11] Li X, Chen H, Lu H, Li W, Chen X, Peng Y, et al. The study on the role of inflammatory cells and mediators in post infectious functional dyspepsia. Scand J Gastroenterol. 2010;45:573–81.
- [12] Suzuki H, Matsuzaki J, Hibi T. What is the difference between Helicobacter pyloriassociated dyspepsia and functional dyspepsia? J Neurogastroenterol Motil. 2011;17:124–30.
- [13] Suzuki H. Post infectious functional dyspepsia A novel disease entity among functional gastrointestinal disorders - Relation to *Helicobacter pylori* infection? J Neurogastroenterol Motil. 2010;16:97–98.
- [14] Kindt S, Tertychnyy A, De Hertogh G, Geboes K, Tack J. Intestinal immune activation in presumed post infectious functional dyspepsia. Neurogastroenterol Motil. 2009;21:832-56.
- [15] Hayat M, Cairns A, Dixon MF, O'Mahony S. Quantitation of intraepithelial lymphocytes in human duodenum: What is normal? J Clin Pathol. 2002;55:393-94
- [16] Lorenzo M, Jeffrey J, Hanina H, Green PH, Rotterdam H, Bhagat G. Duodenal intraepithelial lymphocytosis with normal villous architecture: Common occurrence in *H. pylori* gastritis. Modern Pathology. 2005;18:1134–44.
- [17] Odze RD, Goldblum JR. Surgical Pathology of Gastrointestinal tract, liver, Biliary tract and Pancreas. 2nd ed. Philadelphia: Saunders, Elesvier publication; 2009. Chapter 13, Inflammatory Disorders of the Small Intestine; pp. 327-28.
- [18] Fry L, Seah PP, McMinn RM, Hoffbrand AV. Lymphocytic infiltration of epithelium in diagnosis of gluten-sensitive enteropathy. BMJ. 1972;3:371–74.
- [19] Kakar S, Nehra V, Murray JA, Dayharsh GA, Burgart LJ. Significance of intraepithelial lymphocytosis in small bowel biopsy samples with normal mucosal architecture. Am J Gastroenterol. 2003;98:2027–33.
- [20] Patra R, Chattopadhyay S, De R, Ghosh P, Ganguly M, Chowdhury A, et al. Multiple infection and microdiversity among *Helicobacter pylori* isolates in a single host in India. PLoS One. 2012;7(8)e43370.
- [21] Sodhi JS, Javid G, Zargar SA, Tufail S, Shah A, Khan BA, et al. Prevalence of Helicobacter pylori infection and the effect of its eradication on symptoms of functional dyspepsia in Kashmir, India. J Gastroenterol Hepatol. 2013;28(5):808-13
- [22] Rothenberg ME, Hogan SP. The eosinophil. Annu Rev Immunol. 2006;24:147-74.
- [23] Talley NJ, Walker MM, Aro P, Ronkainen J, Storskrubb T, Hindley LA, et al. Nonulcer dyspepsia and duodenal eosinophilia. An adult endoscopic populationbased case control study. Clin Gastroenterol Hepatol. 2007;5:1175–83.
- [24] Walker MM, Talley NJ, Prabhakar M, Pennaneac'h CJ, Aro P, Ronkainen J, et al. Duodenal mastocytosis, eosinophilia and intraepithelial lymphocytosis as possible disease markers in the irritable bowel syndrome and functional dyspepsia. Aliment Pharmacol Ther. 2009;29:765–73.
- [25] Schappi MG, Borrelli O, Knafelz D, Lindley KJ. Mast cell nerve interactions in children with functional dyspepsia. J Pediatr Gastroenterol Nutr. 2008;47(4):472-80.
- [26] Lin XP, Magnusson J, Ahlstedt S, Dahlman-Höglund A, Hanson L LA, Magnusson O, et al. Local allergic reaction in food-hypersensitive adults despite a lack of systemic food-specific IgE. J Allergy Clin Immunol. 2002;109(5):879-87.
- [27] Samsom M, Verhagen MA, Van Berge Henegouwen GP, Smout AJ. Abnormal clearance of exogenous acid and increased acid sensitivity of the proximal duodenum in dyspeptic patients. Gastroenterology. 1999;116:515–20.
- [28] Barbera R, Feinle C, Read NW. Abnormal sensitivity to duodenal lipid infusion in patients with functional dyspepsia. Eur J Gastroenterol Hepatol. 1995;7:1051– 57.

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